- Peak widths are 50% smaller with HALO® PCS Phenyl-Hexyl
- Impurity peaks are clearly visible with HALO<sup>®</sup> PCS Phenyl-Hexyl since the peak shapes are so sharp, but are not visible at all on the uncharged Phenyl-Hexyl column



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# **INTRODUCTION**

**Example 19 advancedmaterialstechnology** ISC 2024 Liverpool, UK

# **Evaluation of Positively Charged Surface Stationary Phases for Improved**

**Chromatographic Separations of Basic Analytes in Small Molecules and Peptides**

## IMPROVEMENTS TO LC OF BASIC MOLECULES

# HALO**®** PCS STATIONARY PHASES

**Problem:** basic compounds become charged at low pH leading to tailed peak shape as sample load is increased under typical low ionic strength reversed-phase LC and LCMS conditions

- HALO<sup>®</sup> PCS phases improve load tolerance in formic acid vs. traditional uncharged stationary phases making PCS phases useful for analysis of less abundant impurities.
- HALO 160 Å PCS C18 exhibits favorable peak shape for peptides in weakly acidic mobile phase thus expanding the choice of mobile phase for effective LC/MS.
- HALO® PCS phases show symmetrical peak shape for basic, neutral, and acidic analytes.
- All HALO® PCS phases exhibit the speed and resolution advantages of Fused-Core® superficially porous particles.



**Solutions:** To improve peak shape, there are a few options such as adding an ion pair reagent or adding a buffer, but these options are not always 100% compatible with MS detection. Specifically, trifluoroacetic acid (TFA) reduces MS ionization efficiency and phosphate buffer is not MS compatible. Another solution is to use a stationary phase with a positive charged ligand. The HALO® PCS (positive charged surface) product family incorporates a positively charged ligand in addition to a traditional stationary phase on superficially porous silica particles. This stationary phase enables improved peak shape, sample loading, and better impurity analysis.

- 2.7  $\mu$ m particle size with 0.5  $\mu$ m thick shell
- 90 Å pore size for small molecules and 160 Å for peptides and tryptic fragments
- Excellent peak shape and increased loading capacity for basic compounds
- 100% aqueous compatible
- UHPLC and LCMS compatible

• Improved tailing factor and efficiency are obtained with HALO® PCS C18 when compared to a traditional uncharged C18 stationary phase for this mix of 4 tricyclic antidepressants

2.1 x 100 mm, A: Water, 0.1% Formic Acid; B: Acetonitrile, 0.1% Formic Acid; Gradient: 2-35 %B in 10 min.; Flow Rate: 0.3 mL/min; Temperature: 30 °C; Injection: 1.0 µL; Wavelength: PDA, 280 nm

- Gradient separation of 5 variant synthetic peptides + insulin  $B_{ox}$
- Reduced retention time and increased resolution for HALO® PCS C18 Peptide compared to uncharged Peptide C18
- Improved peak widths and reduced tailing in formic acid

### Improved Peak Capacity with HALO**®** PCS C18 Peptide

Improved Impurity Analysis with HALO**®** PCS Phenyl-Hexyl



A: Water, 0.1% Formic Acid; B: Acetonitrile, 0.1% Formic Acid; Gradient: 3-50 %B in 30 min.; Flow Rate: 0.4 mL/min; Temperature: 60 °C; ; Shimadzu NexeraX2 -> diverter valve -> QExactive HF (res=240,000) MarvelXACT Post-Column Plumbing: 50 µm x 350 mm from column to diverter valve 50 µm x 350 mm from diverter valve to union 50 µm x 150 mm from grounding union to HESI I 1.0e92 - tryptic digest trastuzumab 1.25µg/µL ESC18 160Å BH202204 SN2560 2.1x150mm 2.7µm 60°C TIC\_F1 300.00000-2000.00000 m/z 8.q 6.q **Peptide C18** 2.7 µm, 2.1 x 150 mm 4. $\mathsf{q}_{\perp}$ when when I Me i where 2. $\mathsf{q}_{\mathsf{L}}$ 9.Q ... -1.0e8 לן.<mark>4'†® ‡וסף iic digest itcastuzunab 0005 cg/pL 000900 miz 160A BH</mark>2321704 SN1281 2.1x150mm 2.7µm 60°C  $8.0$ **Peak Capacity = 170** 6. $\mathsf{q}$  $4.0$ 2. $\mathbf{\mathfrak{q}}$  ,

 $-1.0200$  4.0 6.0 8.0 10.0 12.0 14.0 16.0 18.0 20.0 22.0 24.2b.0 <u>a di L</u>



- Peak capacities  $(n_{\text{PC}})$  measured with modest load (2  $\mu$ g) of trastuzumab tryptic digest on a 2.1 mm ID column
- $n_{PC}$  based on 12 ID peptides measured using extracted ions (XICs) PW<sub>1/2</sub>, t<sub>R</sub> and  $\Delta t_G$  for this specific sample set
- Decreased peak widths effect notable increase in peak capacity

2.1 x 100 mm, A: Water, 0.1% Formic Acid; B: Acetonitrile, 0.1% Formic Acid; Flow Rate: 0.4 mL/min; Back Pressure: 242 bar; Temperature: 30  $^{\circ}$ C; Injection: 0.5 µL (31 µg) Sample Solvent: 75/25 Water/ACN; Wavelength: PDA, 254 nm, LC System: Shimadzu Nexera X2

- Beta blockers are used for the treatment and/or prevention of heart and circulatory conditions, such as arrhythmias, heart attack, and high blood pressure
- Eleven different beta blockers are separated in under 5 minutes using a HALO® PCS C18 column with UV detection and a mobile phase that is MS compatible

# IMPROVEMENTS TO LC AND LCMS OF PEPTIDES

- The highly efficient 160 Å pore superficially porous particle permits very high throughput analysis
- The example shows separation conducted in less than 2 minutes, with modest backpressure, even at moderate temperature



Bisoprolol (beta-blocker) 2. Bupivacaine (local anesthetic)

#### PEAK IDENTITIES:

2.1 x 100 mm, A: Water, 0.1% Formic Acid; B: Acetonitrile, 0.1% Formic Acid; Isocratic as listed; Flow Rate: 0.4 mL/min; Back Pressure: 206 bar; Temperature: 35 °C; Injection: 1.0 µL Sample Solvent: 90/10 Water/ACN; Wavelength: PDA, 280 nm, LC System: Shimadzu Nexera X2

#### **HALO® PCS C18 Peptide** 2.7 µm, 2.1 x 150 mm

#### **Peak Capacity = 488**

90

110

## Fast Separation of **β-**Blockers

2.1 x 100 mm, A: Water, 0.1% Formic Acid; B: Acetonitrile, 0.1% Formic Acid; Gradient: 3-36% B in 5 min; Flow Rate: 0.4 mL/min; Back Pressure: 281 bar; Temperature: 30 °C; Injection: 1.0 µL Sample Solvent: 93/7 Water/ACN; Wavelength: PDA, 220 nm, LC System: Shimadzu Nexera X2





- HIC isolated mAb with 2 vedotin-ejfv payload conjugated to enfortumab was digested with trypsin
- A single L-chain cystine site was occupied by the payload, with retention identified in the XIC, verified by MS/MS

# **CONCLUSIONS**

## Tryptic digest LCMS analyses to identify payload site for isolated DAR2 ADC



#### High Speed Peptide Analysis

HALO 160 Å PCS C18, 2.1 x 50 mm, A: Water, 0.1% Formic Acid; B: Acetonitrile, 0.1% Formic Acid; Gradient: 0-35 %B in 1.5 min.; Flow Rate: 1.0 mL/min; Temperature: 30 °C; Injection: 1.0 µL; Wavelength: PDA, 280 nm



#### Effect of Organic Solvent on HALO**®** PCS Selectivity

#### Peak Identities:



2.1 x 100 mm, A: Water, 0.1% Formic Acid; B: Specified Solvent, 0.1% Formic Acid; Isocratic at specified % B; Flow Rate: 0.5 mL/min; Temperature: 30 °C; Injection: 1.0 µL Wavelength: PDA, 230 nm, LC System: Shimadzu Nexera X2



- Changing from acetonitrile to methanol gives elution order changes for both PCS C18 and PCS Phenyl-Hexyl
- Increased selectivity and resolution are observed with PCS Phenyl-Hexyl compared to PCS C18 when run with methanol

2.1 x 100 mm, A: Water, 0.1% Formic Acid; B: Acetonitrile, 0.1% Formic Acid; Isocratic as listed; Flow Rate: 0.4 mL/min; Back Pressure: 238 bar; Temperature: 35 °C; Injection: 1.0 µL Sample Solvent: 70/30 Water/ACN; Wavelength: PDA, 254 nm, LC System: Shimadzu Nexera X2



## HALO**®** PCS C18 Compared to Competitor Charged C18



#### • Better tailing and higher efficiency is observed with HALO® PCS C18 for the basic compound (peak 2)

• Peaks 3 & 4 (acids) show symmetrical peak shape with HALO ® PCS C18





- Lowest tailing factor and highest plates found with HALO® PCS Phenyl-Hexyl compared to a charged C18 phase
- Significant improvements are observed when compared to an uncharged Phenyl-Hexyl phase